Protocol for 1.8X bead clean-up

* Adapted from [the Illumina DNA Prep reference guide](https://support.illumina.com/downloads/illumina-dna-prep-reference-guide-1000000025416.html) (step 6b)
* Sent by: Guillaume Dubeaux at Illumina Technical Support
* Purpose: remove contaminants, identified as low 260/230 ratios, prior to library prep
* Uses IPB (Illumina Purification Beads)

Supplies Needed:

* Magnet
* 80% EtOH
* IPB beads
* RSP
* LoBind PCR tubes

Steps

1. Add 1.8X volume of beads to your samples with a low 260/230 ratio (for instance 90 uL of beads for 50 uL of libraries)
2. Pipette 10 times to mix.
3. Incubate at room temperature for 5 minutes.
4. Place on the magnetic stand and wait until the liquid is clear (~5 minutes).
5. Without disturbing the beads, remove and discard the supernatant.
6. Wash two times as follows.
   1. With the plate on the magnetic stand, add 200 μl fresh 80% EtOH without mixing.
   2. Incubate for 30 seconds.
   3. Without disturbing the beads, remove and discard the supernatant.
7. Use a 20 μl pipette to remove and discard residual EtOH.
8. Air- dry on the magnetic stand for 5 minutes.
9. Remove from the magnetic stand.
10. Add 32 μl RSB to the beads (you can use a smaller volume if you want to concentrate your DNA).
11. Pipette to resuspend.
12. Incubate at room temperature for 2 minutes.
13. Place the plate on the magnetic stand and wait until the liquid is clear (~2 minutes).
14. Transfer 30 μl supernatant (or less depending on the volume you added prior) to a new 96- well PCR plate.